Yeast colony PCR

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1. Pick part of a colony into a PCR tube using a sterile pipet tip or toothpick. Mark the location of the picked colony with a dot and number on the bottom of the plate so you can go back and pick the rest of it to grow up for a glycerol stock if it is correct

2. Microwave picked colonies on high for 1 min

3. For each PCR, prepare the following reaction mix:
    - -5 µL 10X Standard Taq buffer (NEB M0273L)
    - -1 µL 10 mM dNTPs (2.5 mM each dNTP)
    - -1 µL 10 µM forward primer
    - -1 µL 10 µM reverse primer
    - -0.5 µL Taq polymerase (NEB M0273L)
    - -41.5 µL H_2O

4. Add 50 µL PCR mix to each microwaved colony

5. Cycle using the following parameters:
    - -95°C, 5’
    - -35 cycles of:
        - -95°C, 30”
        - -55°C, 30”
        - -68°C, 1’/kb
    - -68°C, 5’
    - -Hold at 8°C

6. Add 10 µL 6X orange loading dye (Lifetech R0631), pipet up and down to mix, and load 10 µL on an agarose gel